



6-20-2023

Creation of a hyperplane device for horizontal cellular migration assays

Nicholas M. Bittner

Nelofar Nargis

Ghoulem Ifrene
ghoulem.ifrene@und.edu

Brent Jeffrey Voels

Colin K. Combs
University of North Dakota, colinkelly.combs@UND.edu

[How does access to this work benefit you? Let us know!](#)

Follow this and additional works at: <https://commons.und.edu/bms-pp>



Part of the [Biomedical Engineering and Bioengineering Commons](#)

Recommended Citation

Bittner, Nicholas M.; Nargis, Nelofar; Ifrene, Ghoulem; Voels, Brent Jeffrey; and Combs, Colin K., "Creation of a hyperplane device for horizontal cellular migration assays" (2023). *Biomedical Sciences Posters and Presentations*. 1.

<https://commons.und.edu/bms-pp/1>

This Poster is brought to you for free and open access by the Department of Biomedical Sciences at UND Scholarly Commons. It has been accepted for inclusion in Biomedical Sciences Posters and Presentations by an authorized administrator of UND Scholarly Commons. For more information, please contact und.common@library.und.edu.

Creation of a hyperplane device for horizontal cellular migration assays.

Nicholas M. Bittner¹, Nelofar Nargis¹, Ghoulem Ifrene¹, Brent J. Voels², Colin K. Combs¹

¹Department of Biomedical Sciences, UND School of Medicine & Health Sciences,

²Cankdeska Cikana Community College



Abstract

Cell culture studies routinely seek to monitor cell migration in response to chemoattractant stimuli. Common assays of cell migration employ well inserts and vertical cell migration assessment. This approach does not allow real-time monitoring of cell behavior. To address this need, we sought to develop a horizontal culture platform conducive to time course cell assessment changes in migration, morphology, phenotype etc. Modification of a commercial chamber slide allowed us to quantify cell migration in response to a 20% serum gradient. Based upon this finding, we designed and fabricated a prototype chamber slide for high replicate, real time assessment of cell migration in the serum gradient. The novel chamber slide design was effective for quantifying not only cell migration differences but visualizing cell movement. Optimization of the fabricated design will provide a novel tool for cell biology research.

Methods

The human breast epithelial cell line, MDA-MB-231, from ATCC was grown in 10% serum containing DMEM/F12 media.

Nunc 8 well chamber slides were modified by using a heated 0.8mm needle to bore a channel between wells for horizontal migration assay.

A 3D printer was used to fabricate a novel chamber slide prototype capable of real-time imaging and quantitation of horizontal cell migration.

After 24 hr of migration into serum free or 20% serum containing media wells, cells were fixed with 4% paraformaldehyde, stained using hematoxylin, then counted using an inverted Leica microscope.

Differences in cell migration numbers were calculated using GraphPad software and a Student's t-test.

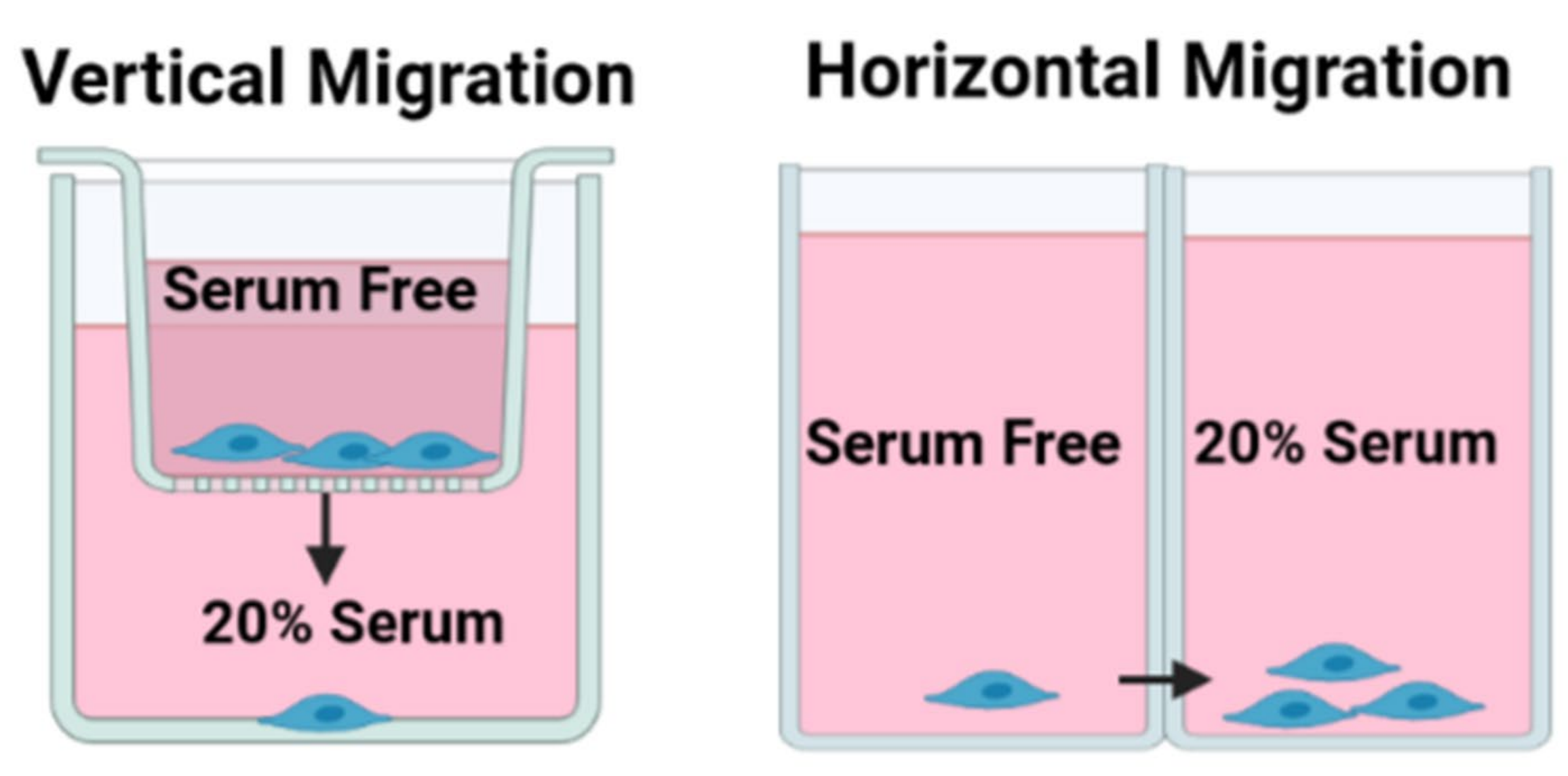


Figure 1. Comparison between vertical and horizontal cell migration assessments.

Results

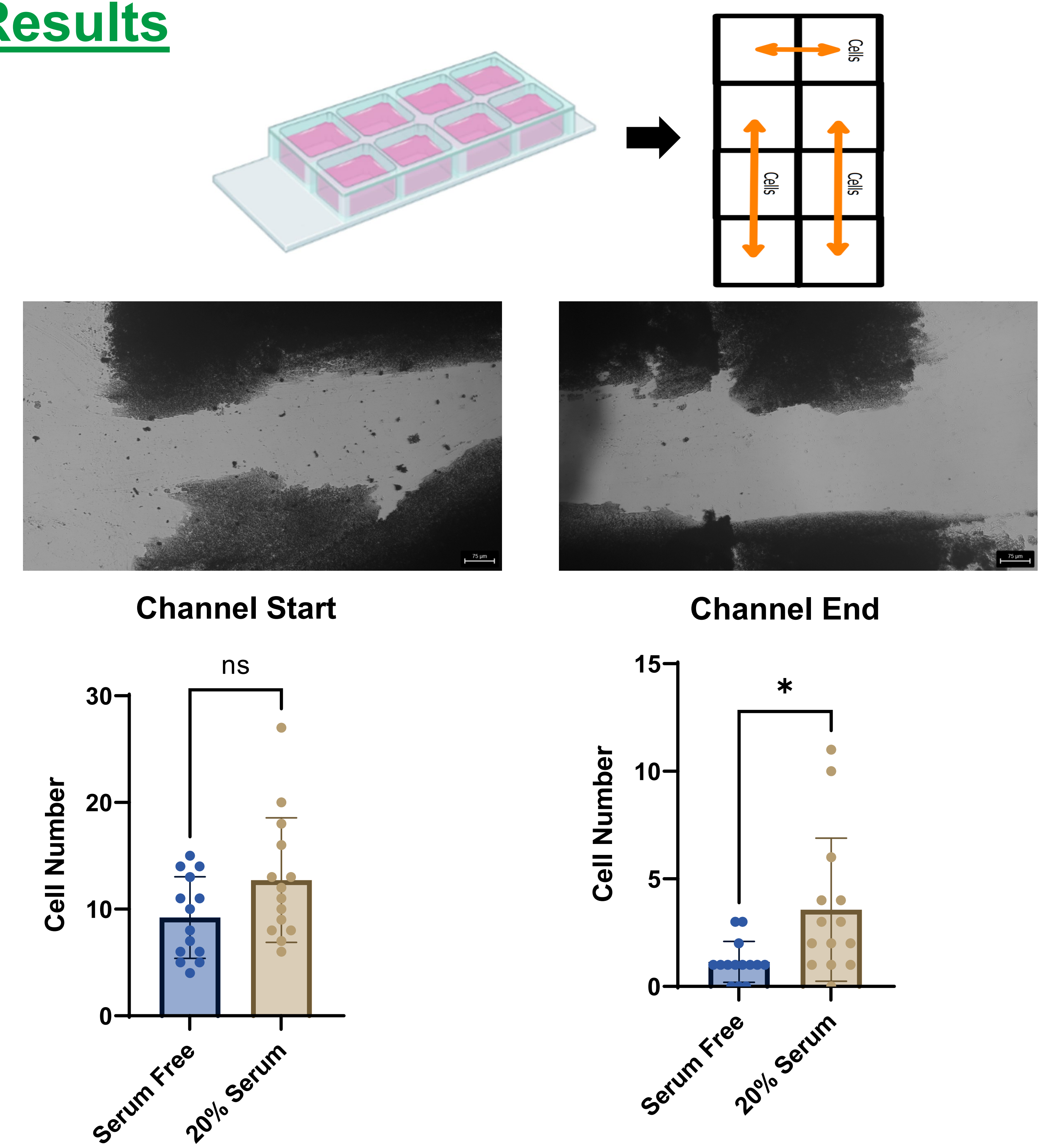


Figure 2. Cell migration in a serum gradient in modified commercial chamber slide. MDA-MB-231 cells were allowed to migrate towards either serum free or 20% serum containing media for 24 hr then cell numbers were counted.

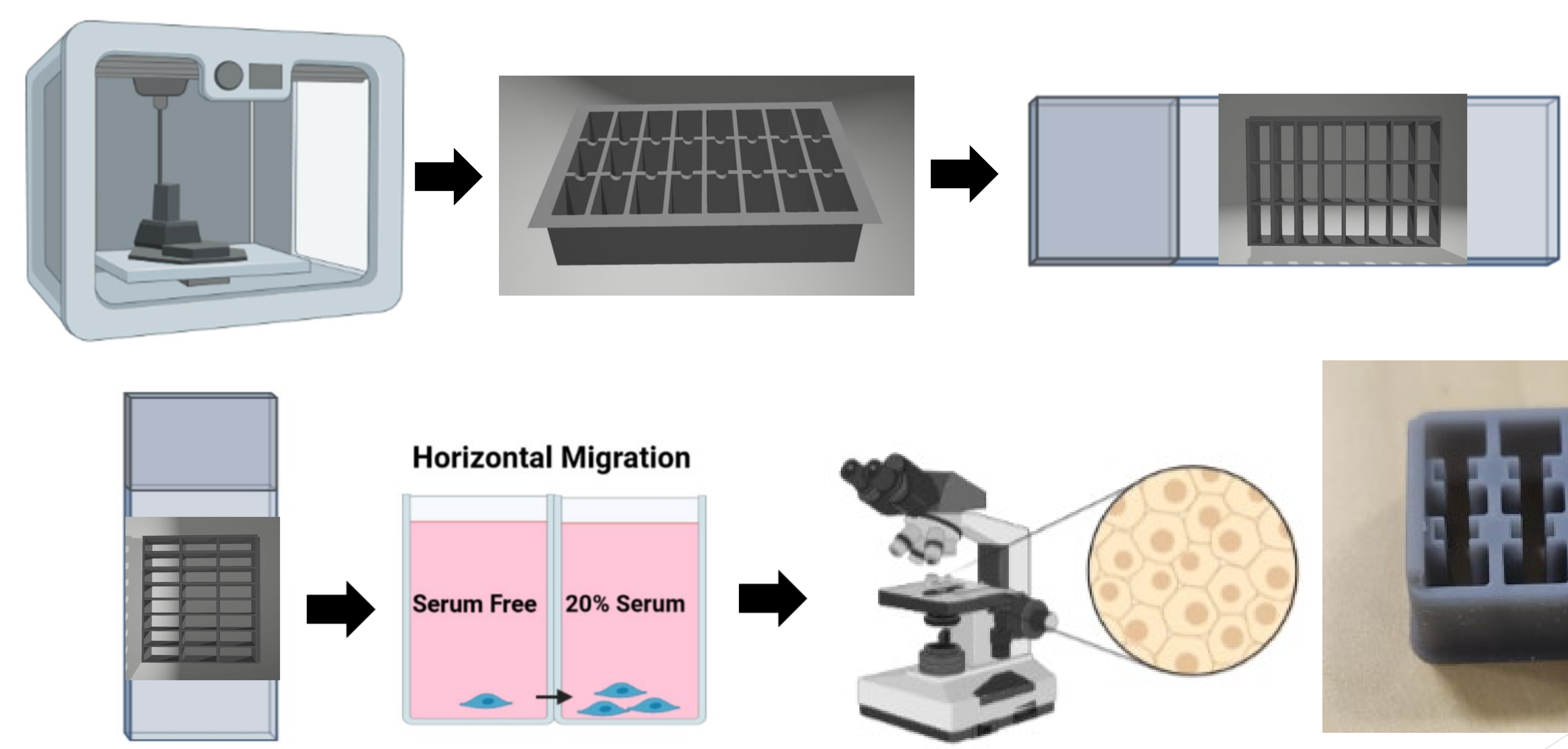


Figure 3. Design of a novel 24 well chamber slide prototype. A prototype chamber slide was fabricated to allow replicates of 8 horizontal migration assays per slide. Current model prototype with slide advancements pictured far right.

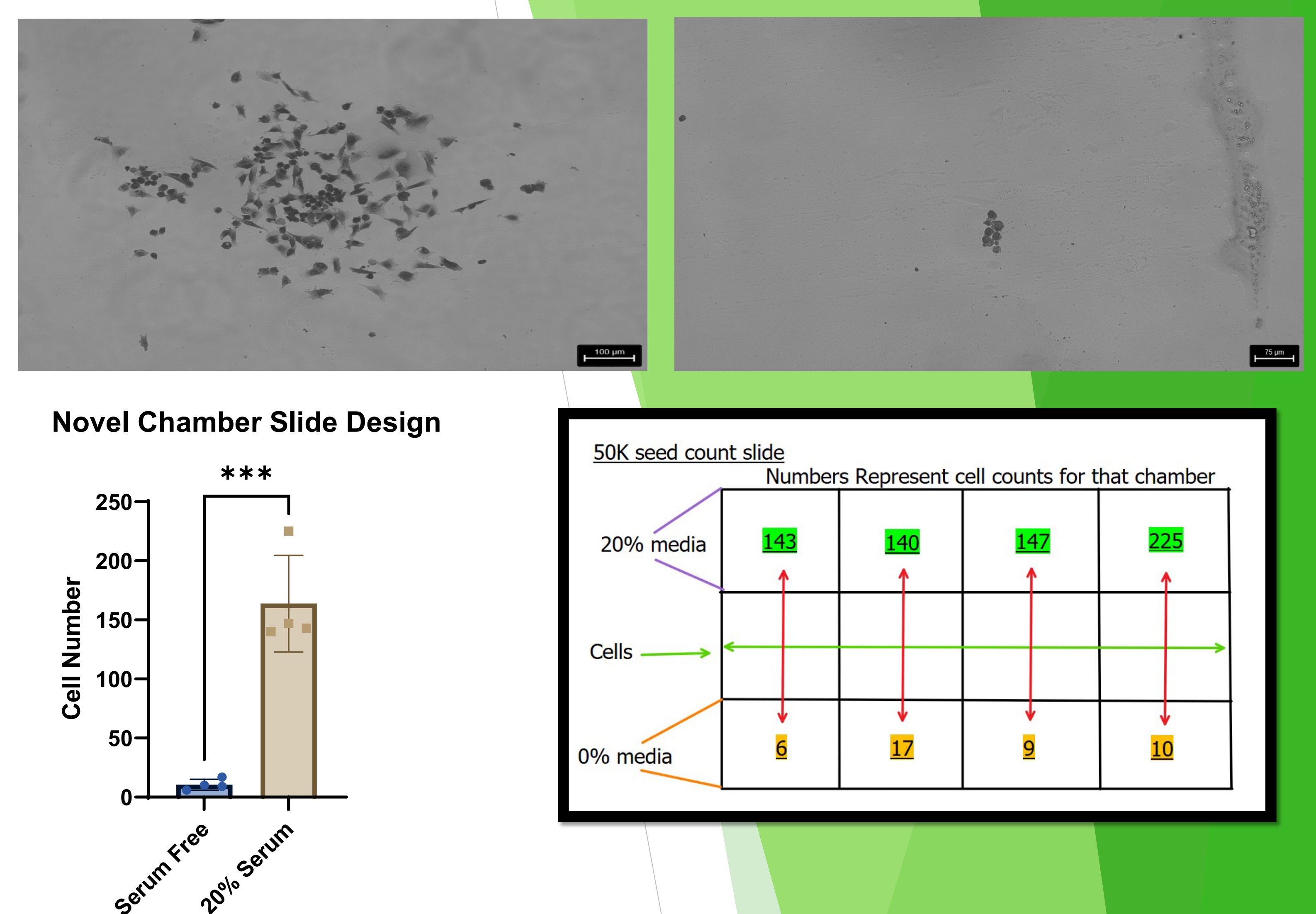


Figure 4. Cell migration in a serum gradient in prototype chamber slide. MDA-MB-231 cells were allowed to migrate towards either serum free or 20% serum containing media for 24 hr then cell numbers were counted.

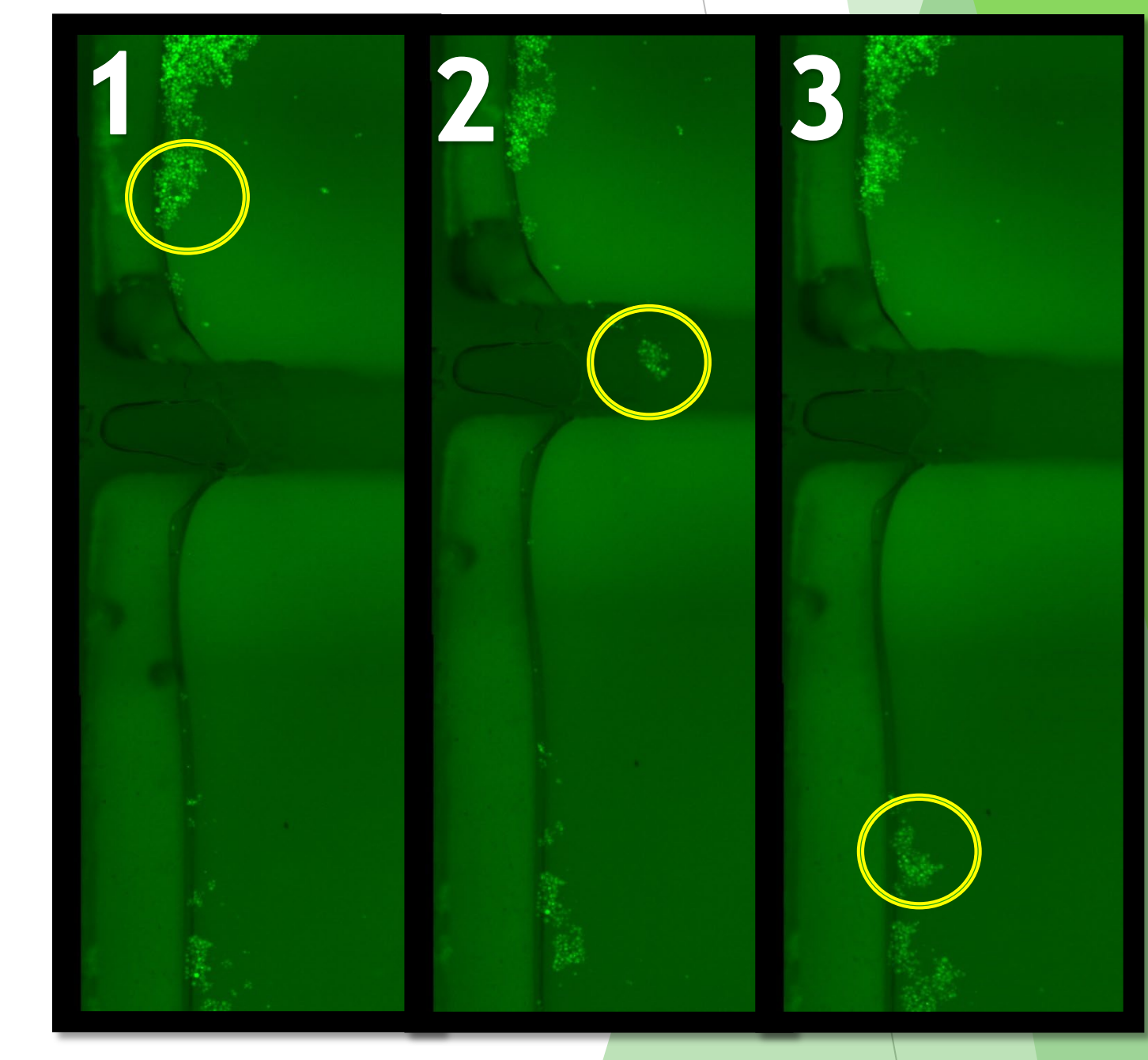


Figure 5. Time-lapse imaging of cell migration. MDA-MB-231 cells were labeled with Cell Tracker Green CMFDA dye and allowed to migrate in the prototype chamber slide for 24 hr, taking pictures every 2 hours. Three consecutive representative images are shown.

Conclusions

- A chamber slide format allows real-time monitoring of cell migration.
- The fabricated, novel chamber slide prototype provides a unique tool for future work focused on cell migration.

Acknowledgments

This research was supported by National Science Foundation Research Experiences for Undergraduates Site grant 1852459, an Institutional Development Award (IDeA) from the National Institute of General Medical Sciences of the National Institutes of Health under grant number P20 GM103442, an NSF RII Track-1 EPSCoR award: ND-ACES award number 1946202, and the Dean of the University of North Dakota School of Medicine & Health Sciences. Images were created using Biorender.com. We thank Joshua Kaelberer for assistance with cell imaging. Imaging studies were conducted in the UND Imaging Core facility supported by NIH grant P20GM113123, DaCCoTA CTR NIH grant U54GM128729, and UNDSMHS funds